ThinPrep® fine needle aspirates (FNA) Quick reference guide

1. Collection.
Collect sample directly into 30ml of CytoLyt® solution. If specimen must be collected in an intravenous solution, use a balanced electrolyte solution.

**Note:** If possible, flush the needle and syringe with a sterile anticoagulant solution prior to aspirating the sample. Some anticoagulants may interfere with other cell processing techniques, so use caution if you plan to use the specimen for other testing.

2. Concentrate by centrifugation.
Centrifuge at 600g for 10 minutes or 1200g for 5 minutes.

3. Pour off supernatant and resuspend cell pellet.
Refer to Procedure A on opposite side of page.

4. Evaluate cell pellet appearance.
Refer to Procedure B on opposite side of page.

5. Add an appropriate amount of specimen (dependent on the size of the cell pellet) to the PreservCyt® solution vial.
Refer to Procedure C on opposite side of page.

6. Allow to stand in PreservCyt solution for 15 minutes.

7. Run on ThinPrep 2000 processor using sequence 2 (FLU/FNA) or ThinPrep 5000 processor using sequence Non-Gyn.
Fix, stain, and evaluate.
Fine needle aspirates

Procedure A

Pour off supernatant and vortex to resuspend cell pellet.

Pour off the supernatant completely to effectively concentrate the sample. To do this, invert the centrifuge tube 180 degrees in one smooth movement, pour off all the supernatant, and then return the tube to its original position as shown in Figure 1. Observe the cell pellet during inversion to avoid accidental loss of cellular material.

Caution: Failure to completely pour off the supernatant may produce a sparse sample and an unsatisfactory slide due to dilution of the cell pellet.

Resuspension can be done on a vortexor or may be achieved by syringing the pellet back and forth with a plastic pipette.

Procedure B

Evaluate cell pellet appearance.

<table>
<thead>
<tr>
<th>Appearance of cell pellet</th>
<th>Procedure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell pellet is white, pale pink, tan or not visible.</td>
<td>Add specimen to PreservCyt® solution vial. See procedure C.</td>
</tr>
<tr>
<td>Cell pellet is distinctly red or brown indicating the presence of blood.</td>
<td>CytoLyt® solution wash.</td>
</tr>
<tr>
<td>Cell pellet is mucoid (not in liquid form). To test for liquid form, draw a small amount of the sample into a pipette and deliver drops back into the tube. If the drops appear stringy or gelatinous, then the mucus must be further liquefied.</td>
<td>CytoLyt solution wash.</td>
</tr>
</tbody>
</table>

Procedure C

Add specimen to PreservCyt solution vial.

Determine the cell pellet size and refer to the table below:

<table>
<thead>
<tr>
<th>Size of cell pellet</th>
<th>Procedure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pellet is clearly visible and the pellet volume is less than 1ml.</td>
<td>Place the centrifuge tube in a vortexor to resuspend the cells in the residual liquid or mix the pellet by syringing it manually with a pipette. Transfer 2 drops of the pellet to a fresh PreservCyt solution vial.</td>
</tr>
<tr>
<td>Pellet is not visible or is scant.</td>
<td>Add the contents of a fresh PreservCyt solution vial (20ml) into the tube. Vortex briefly to mix the solution and pour the entire sample back into the PreservCyt solution vial.</td>
</tr>
<tr>
<td>Pellet volume is greater than 1ml.</td>
<td>Add 1ml of CytoLyt solution into the tube. Vortex briefly to resuspend the pellet. Transfer 1 drop of the specimen to a fresh PreservCyt solution vial.</td>
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</tbody>
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